

# Emergence Of Polymyxin B As A Viable Treatment Option In Comparison To Newer Antimicrobials In Intensive Care Units: A Study In North India

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## ABSTRACT

**Background:** Multidrug resistance (MDR) among gram-negative bacilli has increased substantially limiting the choice of antimicrobials. This study was conducted with the objective to determine the efficacy of tigecycline, polymyxin B, newer fluoroquinolones and newer carbapenems against MDR gram negative isolates.

**Methods:** 90 clinical samples were obtained from ICU patients. On the basis of antibiotic susceptibility to first line antibiotics isolates were divided into 3 groups- a) sensitive to all the first line drugs, b) sensitive only to injectable and c) resistant to all antibiotics except imipenem. These groups were then tested against enoxacin (10 µg), gemifloxacin (5µg), moxifloxacin (5µg), prulifloxacin (5µg), ertapenem (10µg), faropenem (5µg), tigecycline (15µg) and polymyxin B (300 units). Isolates were screened for ESBL, AmpC, CRE and MBL.

**Results:** All the isolates in group 1 were uniformly sensitive to all the new antimicrobials tested. In group 2 susceptibility profile was as follows- 100% sensitive to polymyxin B, 16.6% to tigecycline, 10% to enoxacin, 3.3% to gemifloxacin, moxifloxacin, prulifloxacin, ertapenem and faropenem. In group 3, 81.5% of the isolates were sensitive to polymyxin B, 13.2% to tigecycline, 3.3% each to gemifloxacin and ertapenem.. Isolates of the three groups were uniformly sensitive to imipenem(100%). 2(6.67%) of the isolates were ESBL producers and 30 (33.3%) were AmpC producers. No CRE and MBL were detected.

**Conclusion:** Polymyxin B emerged as most effective antimicrobial in group 2 and group 3 with 100% and 81.5% sensitivity respectively. Use of polymyxin B will prevent injudicious use of imipenem and will decrease escalation of MBLs in our facility.

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## Introduction

The prevalence of multidrug resistance (MDR) among gram-negative bacilli has increased substantially over the years.<sup>[1]</sup>Infections by these MDR gram-negative bacilli often lead to prolonged hospitalization, increased mortality, and greater costs of treatment.<sup>[2]</sup> The emergence and spread of these pathogens in health care settings, has lead to an acute shortage of effective antibiotics which can be effectively used in initial empiric therapy.<sup>[3]</sup>The emergence of extended spectrum  $\beta$  lactamases (ESBL) and AmpC production by Gram negative bacteria further limits the choice of antimicrobials.<sup>[4]</sup>Ultimately carbapenems are used as the drugs of last resort in the treatment of life threatening infections.

Unfortunately in recent years carbapenem resistance is increasingly being reported in Gram negative bacteria.<sup>[5]</sup> Given the alarming state of drug resistance, clearly there is an urgent need for newer antimicrobial agents with novel mechanisms of action to reduce the burden on carbapenems and thus in the process decrease the emergence of carbapenemases.

This study was conducted with the objective to determine the in-vitro activity of tigecycline, polymyxin B, newer fluoroquinolones and newer carbapenems against MDR gram negative isolates.

## Materials and Methods

**Study group:** The study was conducted in the Department of Microbiology on patients admitted to the Intensive Care Unit, Jawaharlal Nehru Medical College and Hospital. The study group comprised of 90 clinical samples from ICU patients obtained from the following sources: surgical site infections [SSI] (70), drains (10), urine (5), tracheal aspirate (3), sputum (1) and Foley's catheter tip (1). Rigorous precautions were taken during sample collection. Clinical significance of the bacteria from tracheal aspirate was assessed as per Shin et al.<sup>[6]</sup> Briefly, sample was rejected if >10 epithelial cells were seen under low power in a direct smear of a gram stained slide. Uncentrifuged urine samples were screened for significant pyuria by direct wet mount. Semiquantitative cultures were put up by using filter paper method.<sup>[7]</sup> Collection and transport was done as per standard protocol.<sup>[7]</sup>

**Processing of sample:** Culture was performed on 5% sheep blood agar, MacConkey agar and BHI broth. Identification was done as per standard guidelines.<sup>[8]</sup>

**Anti-microbial susceptibility testing:** Antibiotic susceptibility testing was performed on Mueller Hinton agar by Kirby Bauer disc diffusion technique as per the CLSI guidelines.<sup>[8]</sup> Bacterial isolates were tested first

against routinely used antibiotics: gentamicin (10 $\mu$ g), amikacin (30 $\mu$ g), amoxicillin (20 $\mu$ g), ceftriaxone (30 $\mu$ g), cefotaxime (30 $\mu$ g), cefoperazone+sulbactam (75/75 $\mu$ g), cefixime (15 $\mu$ g), cefoperazone (75 $\mu$ g), ceftazidime (30 $\mu$ g), ofloxacin (5 $\mu$ g), piperacillin (100 $\mu$ g), piperacillin-tazobactam (100/10 $\mu$ g), tobramycin (10 $\mu$ g) and imipenem (10 $\mu$ g). Antimicrobial susceptibility controls used were *Escherichia coli* ATCC 25922 and *Pseudomonas aeruginosa* 27853.

Using this set of antibiotics, ESBLs, AmpCs and CREs were detected as follows:

**Detection of extended spectrum beta lactamases :** Screening of possible ESBL production was done by using ceftriaxone (30 $\mu$ g) and cefoperazone (75 $\mu$ g). Those isolates with zone diameters less than 25mm for ceftriaxone and less than 22mm for cefoperazone were subsequently confirmed for ESBL production. Confirmation was done by noting the potentiation of the activity of cefoperazone in the presence of cefoperazone sulbactam.<sup>[8]</sup>

**Detection of inducible and derepressed AmpC beta lactamase:** Detection of AmpC beta lactamase was done on isolates resistant to ceftriaxone (30 $\mu$ g), cefixime (15 $\mu$ g), cefoperazone (75 $\mu$ g) and cefoperazone sulbactam (75/75 $\mu$ g). Induction of AmpC synthesis was based on the disc approximation assay using imipenem as inducer.<sup>[8]</sup>

**Detection of CRE (Carbapenem resistant Enterobacteriaceae):** Isolates demonstrating zone sizes of less than 16 mm around imipenem were identified as CRE.

**Detection of Metallo-beta-lactamases:** MBL were detected by modified Hodge test and Double Disc synergy test using EDTA.<sup>[9]</sup>

**Study groups:** On the basis of susceptibility profile to first line drugs and drug resistance markers, the isolates were divided into 3 groups:

**Group 1-** 30 bacterial isolates susceptible to all the routinely used/ tested antibiotics.

**Group 2-** 30 bacterial isolates resistant to all the routinely tested antibiotics except to injectable drugs (amikacin, gentamicin, cefoperazone+sulbactam, piperacillin+tazobactam, tobramycin). This group contained ESBL producers.

**Group 3-** 30 bacterial isolates resistant to all drugs except imipenem. This group consisted of AmpC producers.

**Susceptibility to newer antimicrobials:** These groups were further tested against enoxacin (10  $\mu$ g), gemifloxacin (5 $\mu$ g), moxifloxacin (5 $\mu$ g), prulifloxacin (5 $\mu$ g), ertapenem (10  $\mu$ g), faropenem (5 $\mu$ g), tigecycline (15 $\mu$ g)

and polymyxin B (300 units). All discs were obtained from HiMedia, India.

## Result

The organisms isolated were *Escherichia coli* (n=45), *Klebsiella pneumoniae*(n=20), *Citrobacter species* (n=15), *Serratia species* (n=5), *Acinetobacter species* (n=4) and *Proteus mirabilis* (n=1).. Amongst them 2 (6.67%) of the isolates were ESBL producers and 30(33.3%) were AmpC producers. Table 1 shows the susceptibility pattern to injectable antibiotics in group 2. All the isolates were susceptible to amikacin (100%), 3(10%) to gentamicin, 2(6.67%) each to cefoperazone/sulbactam, tobramycin and piperacillin/tazobactam. No MBL and CRE were detected. Table 2 shows compares susceptibility of the three groups to newer antimicrobials. All the isolates in group 1 were uniformly sensitive to all the routine and the newer antimicrobials tested. In group 2 which also contained ESBLs, susceptibility profile was as follows-100% sensitivity was observed to polymyxin B, 16.6% to tigecycline, 10% to enoxacin, 3.3% to gemifloxacin, moxifloxacin, prulifloxacin, ertapenem and faropenem. In group 3, 81.5% of the isolates were sensitive to polymyxin B, 13.2% to tigecycline, 3.3% each to gemifloxacin and ertapenem. All the isolates were resistant to moxifloxacin,

**Table 1-Susceptibility profile of pathogens to Injectable Antibiotics in Group 2**

Antibiotics	NO. of isolates (n=30)
Amikacin,	30(100%)
Gentamicin	3(10%)
Cefoperazone+sulbactam	2(6.67%)
Piperacillin+tazobactam	2(6.67%)
Tobramycin	2(6.67%)

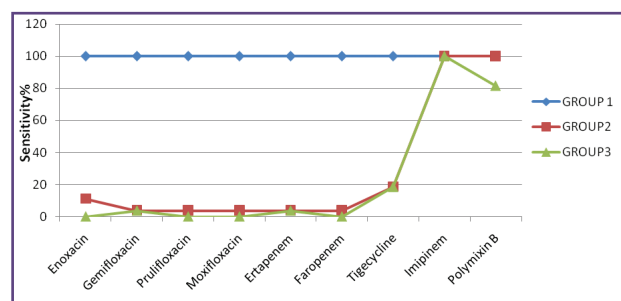
\*Maximum isolates were sensitive to Amikacin (100%)

**Table 2- Comparison of Susceptibility of the Three Groups to Newer Antimicrobials**

Antibiotics	Group 1(n=30)	Group 2(n=30)	Group 3(n=30)
Enoxacin	30(100%)	3(10%)	0(0%)
Gemifloxacin	30(100%)	1(3.3%)	1(3.3%)
Moxifloxacin	30(100%)	1(3.3%)	0(0%)
Prulifloxacin	30(100%)	1(3.3%)	0(0%)
Ertapenem	30(100%)	1(3.3%)	1(3.3%)
Faropenem	30(100%)	1(3.3%)	0(0%)
Tigecycline	30(100%)	5(16.6%)	4(13.2%)
Polymyxin B	30(100%)	30(100%)	24(81.5%)

\* Tigecycline and Polymyxin B showed better results in group 2 as compared to group 3

prulifloxacin, enoxacin and faropenem. Group 3 isolates showed high level of resistance to both aminoglycosides and fluoroquinolones. Isolates of the three groups were uniformly sensitive to imipenem (100%). Figure 1 shows trend of antimicrobial sensitivity in different groups. Barring imipenem, only polymyxin B followed by tigecycline demonstrated encouraging results. Both polymyxin B and tigecycline worked better in group 2 than in group 3.



**Fig. 1: Trend of antimicrobial sensitivity in different groups**

## Discussion

Increasing bacterial resistance to the commonly used antimicrobial agents is increasing and is a matter of grave public health concern, particularly in patients with serious and complicated nosocomial infections. The emergence of ESBL and AmpCs, not to mention the MBLs has led to severely limited therapeutic options, resulting in increased morbidity and mortality.

In this study, prevalence of ESBL was 6.67% while AmpC was much higher at 33.3%. In other studies, AmpC levels were usually lower than ESBLs.<sup>[10,11,12]</sup> The elevated levels of AmpC is alarming as the usage of imipenem increases accordingly.

Polymyxin B emerged as the most effective antimicrobial in group 2 and group 3 with 100% and 81.5% sensitivity respectively. The result was similar to the study done by Castanheira who reported 88.1% of CRE isolates were susceptible to Polymyxin B.<sup>[13]</sup> There has been resurgence in the use of polymyxins as the drugs of last resort for the treatment of infections caused by MDR gram negative pathogens which are resistant to all other currently available antibiotics. Polymyxin B, a polypeptide cationic antibiotic is active against a variety of gram negative bacilli, including most clinically relevant enterobacteriaceae. It is rapidly acting bacteriocidal agent with dose adjustments required for patients with renal impairment, including decreasing daily dose and extending administration intervals.<sup>[14]</sup>

In our study, only 16.6% of the bacterial isolates in group 2 and 13.2% of the isolates in group 3 showed susceptibility

to tigecycline but it had a better susceptibility profile than other newer antimicrobials tested including ertapenem and faropenem. Other studies however have reported good activity.<sup>[15,16,17]</sup> Tigecycline, a newer semi-synthetic glycylicycline derived from minocycline is a promising molecule in the treatment of infections caused by MDR organisms. It is a bacteriostatic agent and has potent invitro activity against several bacteria including ESBL producing Enterobacteriaceae and carbapenem resistant *Acinetobacter* spp. Furthermore, it is unaffected by the known mechanisms of resistance to tetracycline and minocycline such as efflux pumps and ribosomal protective mechanisms.

Although ertapenem is approved for complicated intra-abdominal infections, complicated skin and skin structure infections, community acquired pneumonia, complicated urinary tract infections including pyelonephritis due to susceptible pathogens, and acute pelvic infections, we observed an unexpectedly low sensitivity of 3.7% for ertapenem in our study. This is in sharp contrast to other studies which reported that ertapenem was strongly active against ESBL and AmpC producing gram negative bacteria.<sup>[18,19]</sup> As there is a need for new oral options for treatment of multidrug resistant gram negative bacteria, we also evaluated the in vitro activity of faropenem, an oral penem. But again resistance ranging from 96.3% to 100% was observed. Other studies have shown better activity of faropenem against MDR bacteria.<sup>[20,21]</sup>

The newer fluoroquinolones like enoxacin, prulifloxacin, gemifloxacin and moxifloxacin have broad-spectrum bactericidal activity, excellent oral bioavailability, good tissue penetration and favorable safety and tolerability profiles. This is the first study which evaluated the role of newer fluoroquinolones moxifloxacin, prulifloxacin, gemifloxacin and enoxacin in MDR gram negative bacteria from India. However poor results were elicited with low sensitivity (0%-11.1%). Enoxacin was active against 3.7% isolates in group 2 patients. In group 3, the picture was even more dismal.

## Conclusion

After assessing 8 drugs of four antimicrobial groups, we recommend Polymyxin B as empiric treatment in seriously ill patients.

## Funding

None

## Competing Interests

None declared

## References

1. Nordmann P, Cuzon G, Naas T. The real threat of Klebsiella pneumonia carbapenemase producing bacteria. *Lancet Infect. Dis.* 2009;9:228–236.
2. Lautenbach E, Patel JB, Bilker WB, Edelstein PH, Fishman NO. Extended- spectrum b-lactamase-producing *Escherichia coli* and *Klebsiella pneumoniae*: risk factors for infection and impact of resistance on outcomes. *Clin Infect Dis* 2001; 32:1162–1171.
3. Reinert RR, Low DE, Rossi F, Zhang X, Wattal C, Dowzicky MJ. Antimicrobial susceptibility among organisms from the Asia/Pacific Rim, Europe and Latin and North America collected as part of TEST and the invitro activity of tigecycline. *J Antimicrob Chemother* 2007;60: 1018-1029.
4. Rizvi M, Khan F, Shukla I, Malik A, Shaheen. Rising Prevalence of Antimicrobial Resistance in UTI During Pregnancy: Necessity for Exploring Newer Treatment Options. *Journal of Laboratory Physicians* 2011;3:98-103.
5. Pramod MS, Robin DI. Ertapenem, the first of a new group of carbapenems. *J. Antimicrob. Chemother.* 2003; 52: 538-542.
6. Shin YM, Oh YM, Kim MN, Shim TS, Lim CM. Usefulness of Quantitative Endotracheal Aspirate Cultures in Intensive Care Unit Patients with Suspected Pneumonia. *Korean Med Sci* 2011; 26: 865-869.
7. Collee JG, Fraser AG, Marmion BP, Simmons A. Mackey and McCartney practical Medical Microbiology. In: Collee JG, Miles RS, Watt B, ed: Tests for the identification of Bacteria. 14th ed. New Delhi, India: Elsevier, 2006:131-149.
8. Clinical and Laboratory Standards Institute 2014.. Performance standards for antimicrobial susceptibility testing: twenty fourth informational supplement: Approved standards M100-S24. Clinical and Laboratory Standards Institute, Baltimore, USA. 2014.
9. Lee KY, Chong HB, Shin YA, Yong KD, Yum JH. Modified Hodge test and EDTA disc synergy tests to screen metallo beta lactamase producing strains of *Pseudomonas* and *Acinetobacter* species. *Clin Microbiol Infect* 2001; 7:88-91.
10. Singh RK, Pal NK, Banerjee M, Sarkar S, Gupta MS. Surveillance on extended spectrum  $\beta$  lactamase and ampc  $\beta$  lactamase producing gram negative isolates from nosocomial infections *Archives of Clinical Microb.* 2012;3:1-7.
11. Grover N, Sahni AK, Bhattacharya S. Therapeutic challenges of ESBLs and AmpC beta-lactamase

- producers in a tertiary care center. *Med. J armed forces, India*, 2013;69: 4 -1 0.
12. Singhal S, Mathur T, Khan S, Upadhayay DJ, Chugh S. Evaluation of methods for Amp C  $\beta$  lactamase in gram negative clinical isolates from tertiary care hospitals. *Indian J of Med. Microb.* 2005;3:120-124.
  13. Castanheira M, Sader HS, Deshpande LM, Fritsche TR, Jones RN. Antimicrobial activities of tigecycline and other broad-spectrum antimicrobials tested against serine carbapenemase and metallo- $\beta$ -lactamase-producing Enterobacteriaceae: report from the SENTRY Antimicrobial Surveillance Program. *Antimicrob Agents Chemother* 2008;52:570–573.
  14. Argyris M, Matthew EF. Colistin and Polymyxin B in Critical Care. *Crit Care Clinics* 2008;24(2):377–391.
  15. Sekar M, Sekar U. Tigecycline Against Gram Positive and Gram Negative Isolates in a Tertiary Care Hospital. *J of Clin and Diagnos Research*, 2011;5(8): 1559-1563.
  16. Behera B, Das A, Mathur P, Kapil A, Gadepalli R, Dhawan B. Tigecycline susceptibility report from an Indian tertiary care hospital. *Indian J Med Res* 2009;129:446-450.
  17. Manoharan A, Chatterjee S, Madhan S, Mathai D. Evaluation of tigecycline activity in clinical isolates among Indian medical centers. *Indian J Pathol Microbiol* 2010;53 (4):734-737.
  18. Livermore DM, Oakton KJ, Michael W C, Warner M. Activity of Ertapenem (MK-0826) versus Enterobacteriaceae with Potent  $\beta$  Lactamases. *Antimicrobial agents and chemother.* 2001;45:2831–2837.
  19. Lee NY, Lee CC, Huang WH, Tsui KC, Hsueh PR, Ko WC. Carbapenem therapy for bacteremia due to extended-spectrum- $\beta$ -lactamase-producing *Escherichia coli* or *Klebsiella pneumoniae*: implications of ertapenem susceptibility. *Antimicrob Agents Chemother.* 2012 ;56:2888-2893.
  20. Mushtaq S, Hope R, Warner M, David M. Livermore Activity of faropenem against cephalosporin-resistant Enterobacteriaceae. *Journal of Antimicrobial Chemotherapy* 2007; 59:1025–1030.
  21. Livermore DM, Mushtaq S, Nguyen T, Warner M. Strategies to overcome extended-spectrum  $\beta$ -lactamases (ESBLs) and AmpC  $\beta$ -lactamases in shigellae. *Int J Antimicrob Agents*, 2011;37(5):405-409.